

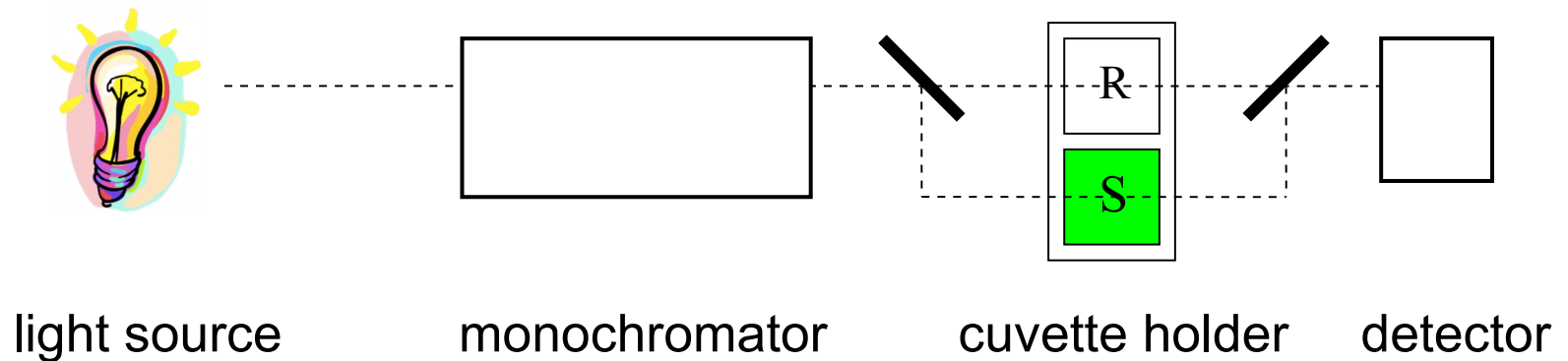
Spectroscopy

Spectroscopy

Absorption

Reported by: 1) percent transmission ($\%T_\lambda = 100 \times I_\lambda / I_{0,\lambda}$)
2) absorbance ($A_\lambda = \log \{I_{0,\lambda} / I_\lambda\} = \epsilon_\lambda c l$)

Measured by: single or double beam spectrophotometer



Spectroscopy

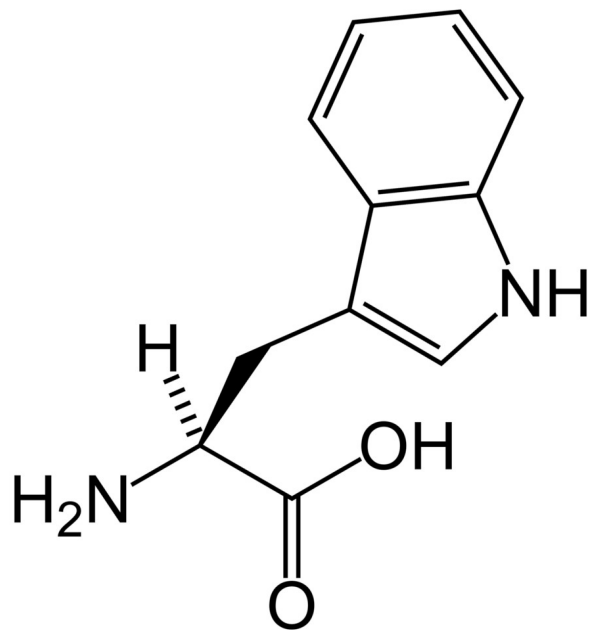
Significance of deviations from the Beer-Lambert Law:

If nothing happens in a solution as a function of concentration of the component solutes, that is there are no concentration-dependent interactions, then the OD should increase linearly with increases in concentration of the components (this requires that all solute components have proportional increases in concentration or that different components have equivalent extinction coefficients).

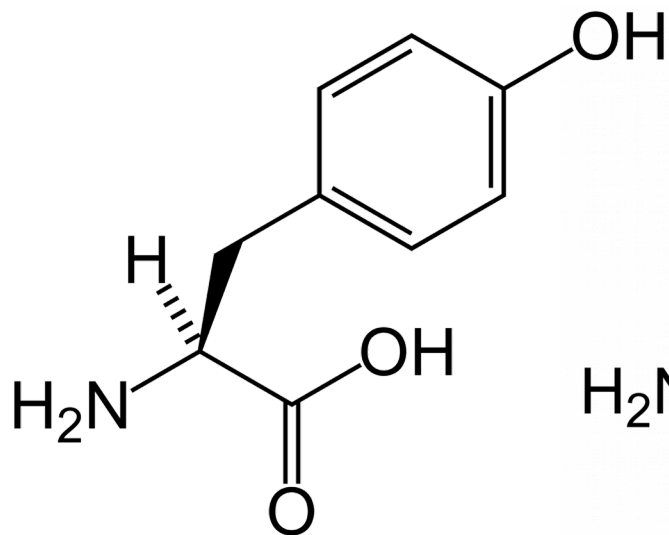
Deviations mean interactions. The change in absorbance versus concentration will follow the thermodynamics of the system, reflecting cooperativity, positive or negative, or lack thereof.

Spectroscopy

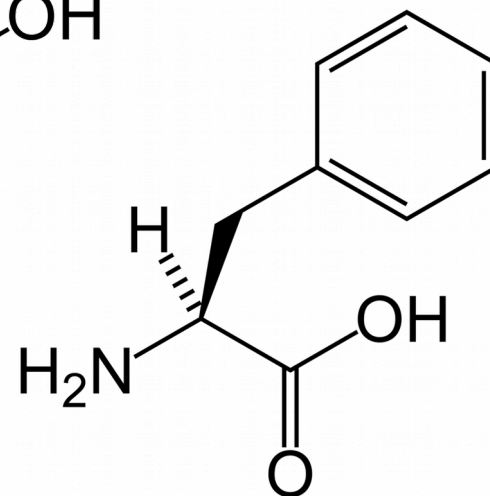
Aromatic Amino Acids



Trp (W)



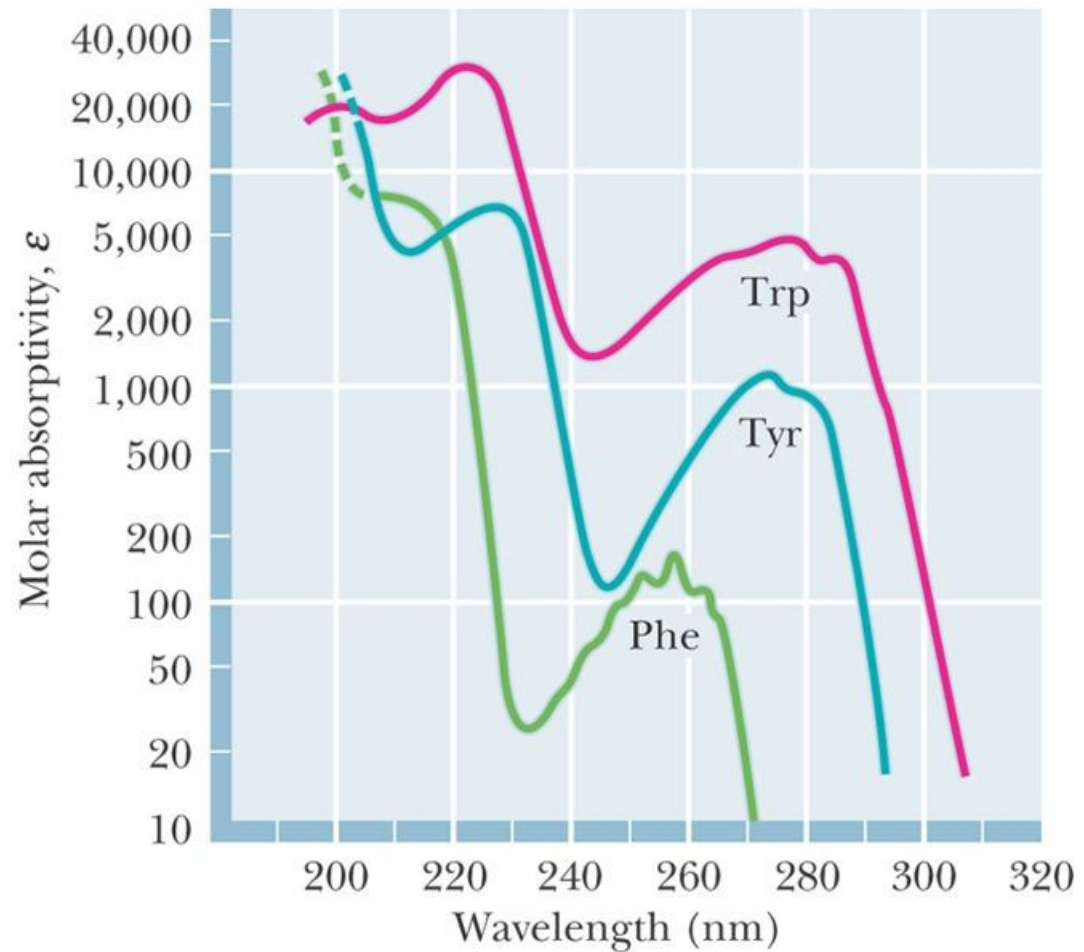
Tyr (Y)



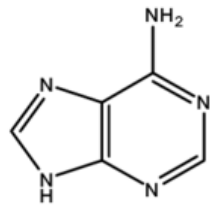
Phe (F)

Spectroscopy

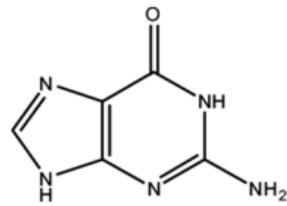
The ultraviolet absorption spectra of the aromatic amino acids at pH 6. (From Wetlaufer, D.B., 1962. *Ultraviolet spectra of proteins and amino acids*. *Advances in Protein Chemistry* **17**:303–390.)



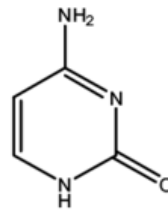
Spectroscopy



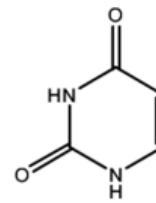
Adenine
A



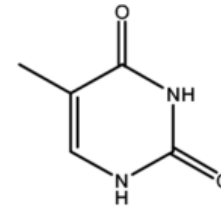
Guanine
G



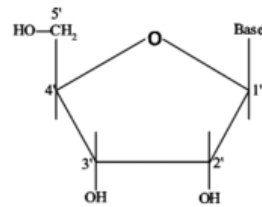
Cytosine
C



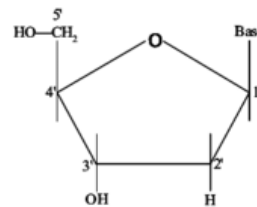
Uracil
U



Thymine
T



Ribose

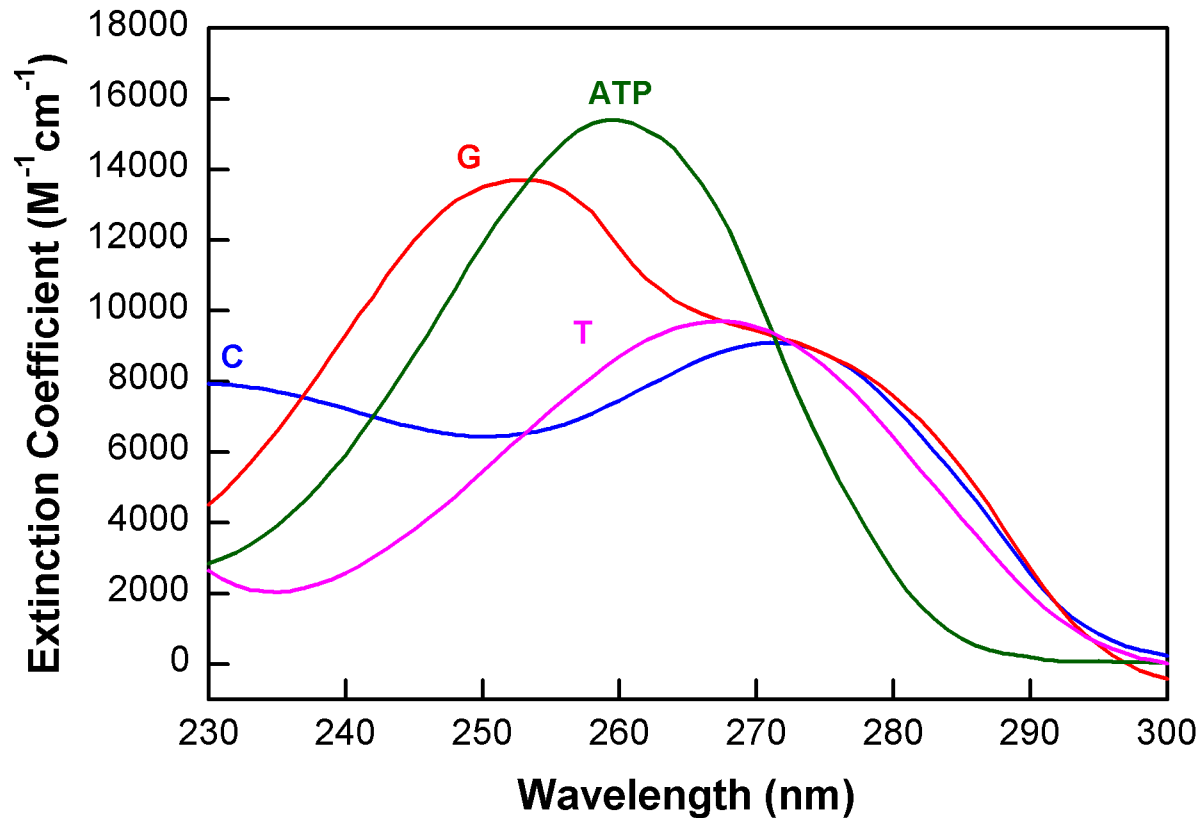


Deoxyribose

Figure 1.1. Structures of nucleic acid constituents

Spectroscopy

Absorption Spectra of the Nucleic Acids



Spectroscopy

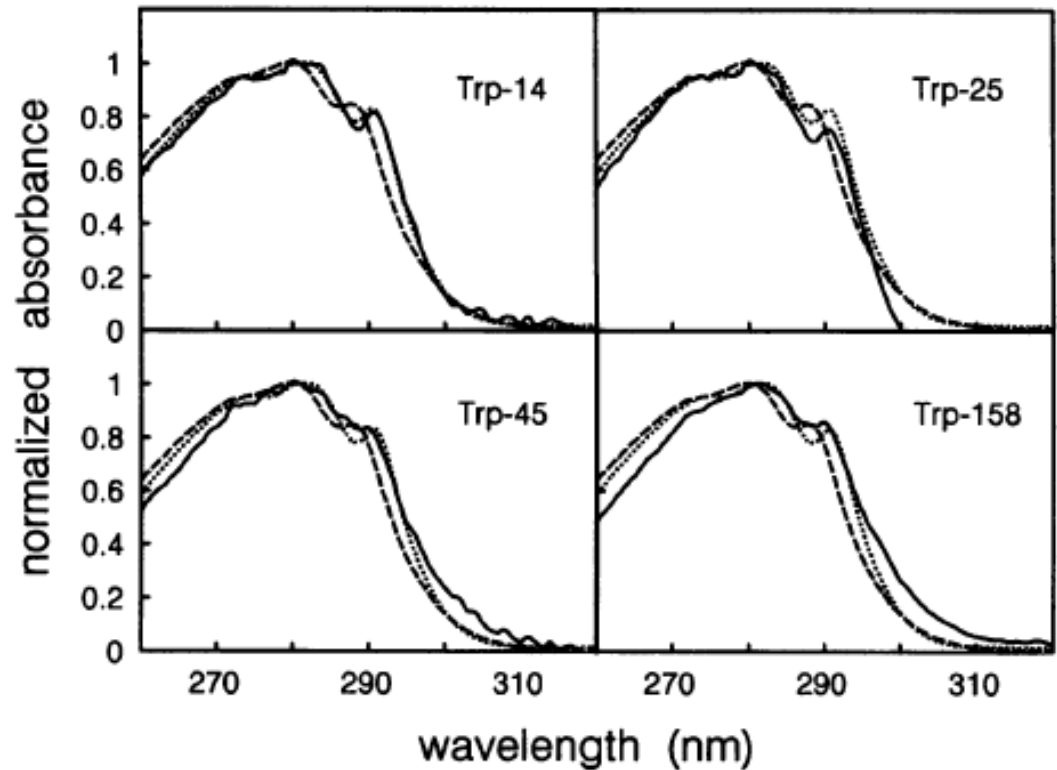
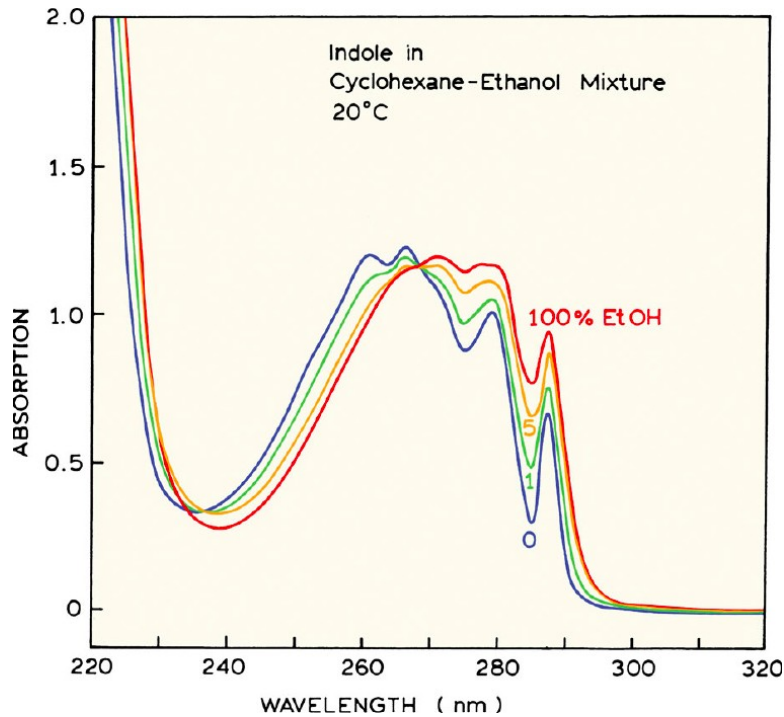
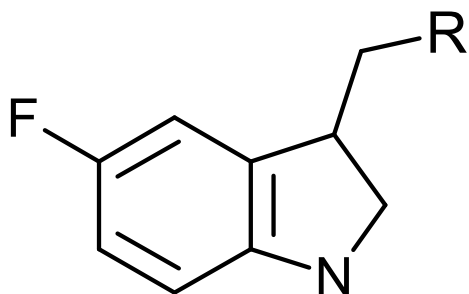


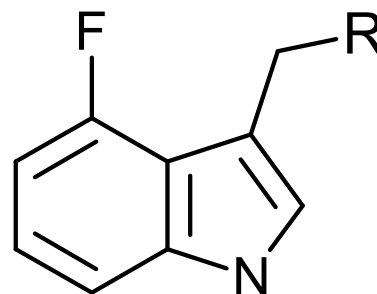
FIGURE 4 Comparison of normalized absorbance spectra of the four Trp in sTF (—) with that of NATA in aqueous buffer (---) and in dioxane (....).

Spectroscopy

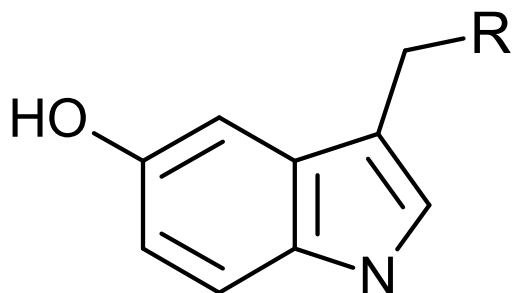
Tryptophan Analogs



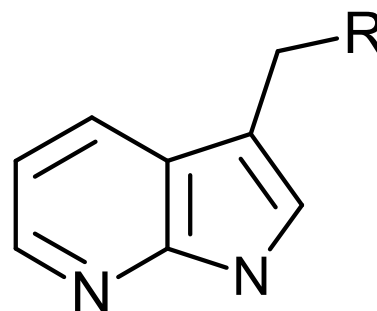
5-fluorotryptophan



4-fluorotryptophan



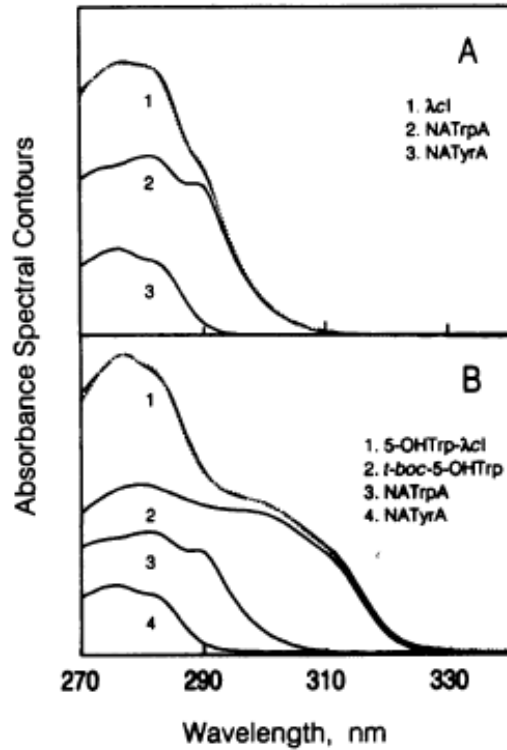
5-hydroxytryptophan



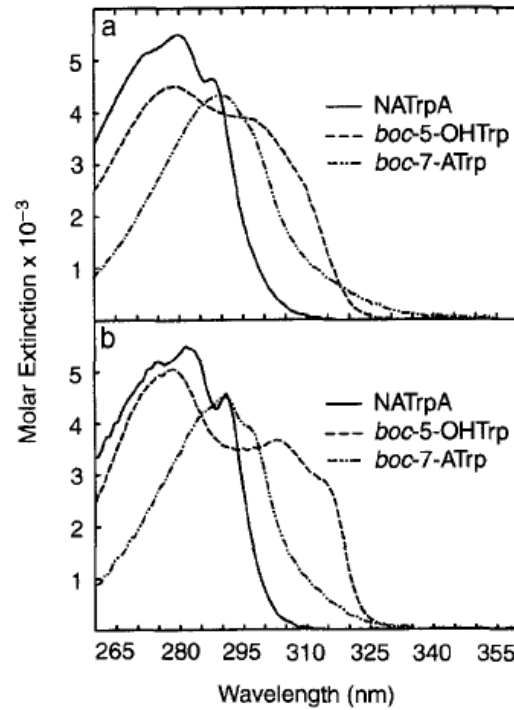
7-azatryptophan

Spectroscopy

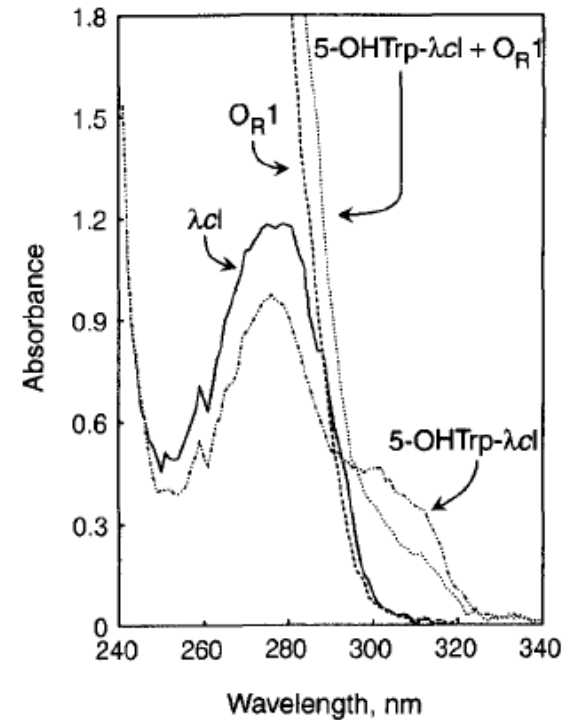
Tryptophan Analogs



Ross et al., Proc Natl Acad Sci USA 89, 12923 (1992)



Ross et al., Meth Enzymol 278, 151 (1998)



Ross et al., Meth Enzymol 278, 151 (1998)

Spectroscopy

Tryptophan Analogs

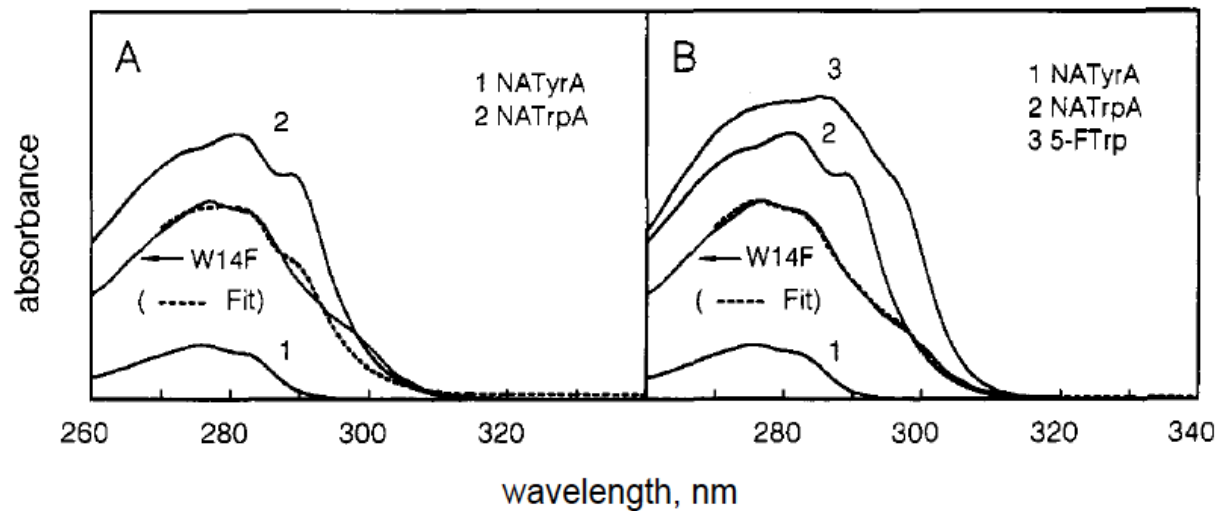
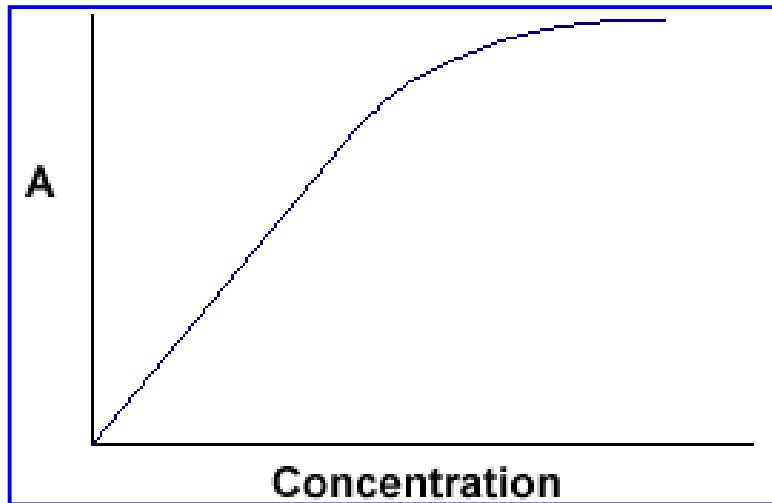


Figure 2.2. LINC analysis of W14F sTF expressed in the presence of 5-FTrp. Panel A shows the fit from 270 to 340nm (dashed line) of the protein absorbance spectrum (solid line) using the NATyrA and NATrpA basis sets. Panel B shows the corresponding fit (dashed line) when 5-FTrp is included as a third basis set.

Spectroscopy

Relationship between Absorbance and Concentration



Why is this plot not linear for the entire range?

At some point the absorbance is so high that not sufficient light passes through to the detector, and linearity is no longer satisfied.

The absorbance at which an instrument becomes non-linear depends on the following factors:

1. Concentration of the analyte
2. Lamp intensity at the measured wavelength
3. Extinction coefficient of the analyte at measured wavelength
4. Sensitivity of detector at the measured wavelength

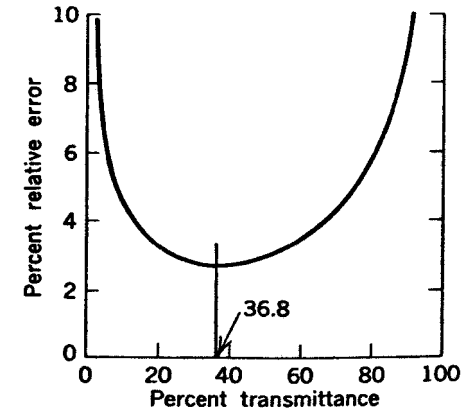
• **For best accuracy, always measure between 0.1 – 1.0 OD**

Spectroscopy

- Concentration

Beer-Lambert Law (Beer's Law): $A_{\lambda} = \epsilon_{\lambda} c l$

most accurate range to obtain c : between 15 and 65 %T
which is A between 0.2 and 0.8



- Identification of Chromophore(s) by Spectra

Are spectra from multiple chromophores in a macromolecule additive?

$$A(\lambda, \text{sample}) = \alpha A(\lambda, a) + \beta A(\lambda, b) + \gamma A(\lambda, c) + \delta A(\lambda, d) + \dots$$

LINCS: LINear Combination of Spectra

approximately linear in proteins

but not linear in DNA or RNA

- Investigate Interacting Systems by Difference Spectra

Quantification of Binding

Evaluation of Conformational Changes

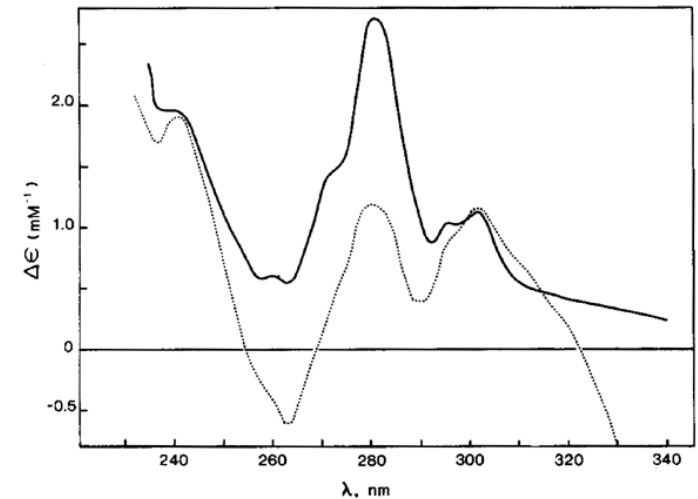


FIG. 2. Absorption difference spectra due to ternary complex formation with liver alcohol dehydrogenase: —, enzyme·NAD⁺·tri-fluoroethanol complex; ····, enzyme·NADH·isobutyramide complex.

Laws and Shore, J. Biol Chem 254, 2582 (1979)

Spectroscopy

PRINCIPLES OF FLUORESCENCE SPECTROSCOPY



Sir George Stokes
1819-1903

Born in County Sligo, Ireland, Stokes was the Lucasian Professor of Mathematics at Cambridge from 1849 until his death in 1903.

Some major contributions:

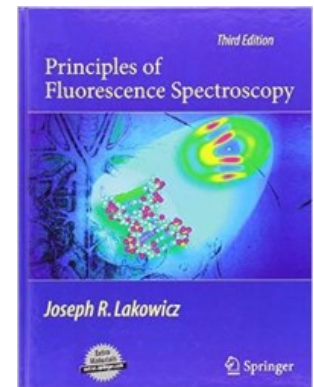
Fluid dynamics (Stokes' Law)

Wave theory of light

Polarization of light

Fluorescence of minerals

Stokes' line (Raman scatter)



Spectroscopy

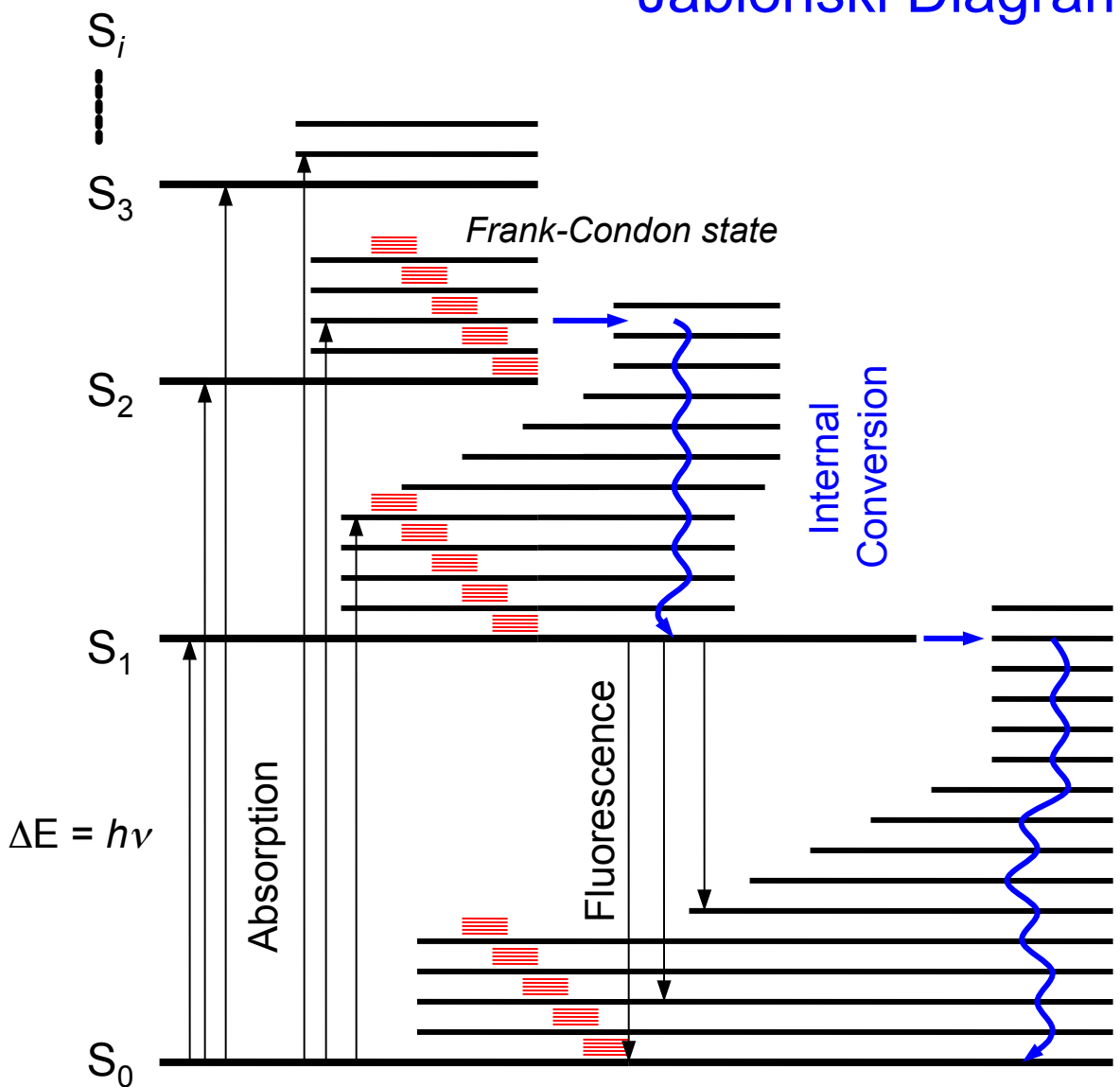
Jablonski Diagram

Time Scales

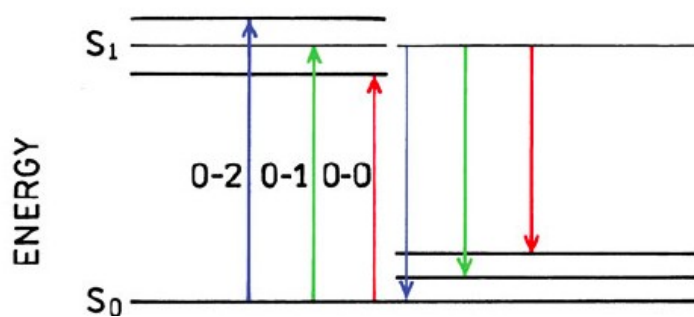
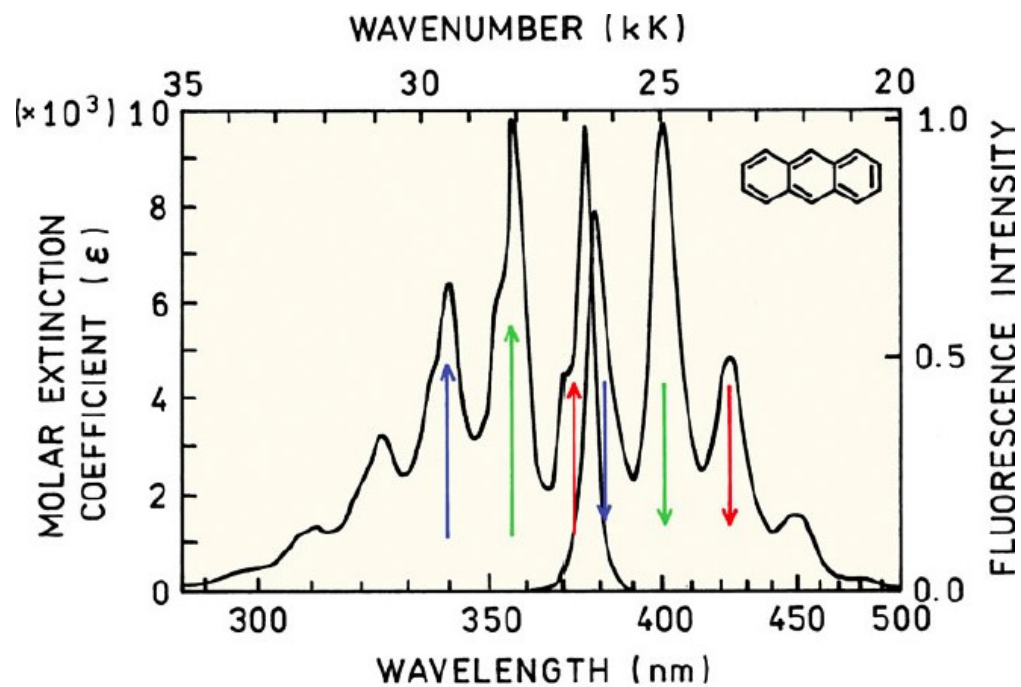
Abs.: $\sim 10^{-15}$ s

IC.: $< 10^{-12}$ s

Fluor.: $\sim 10^{-9}$ s



Spectroscopy



Spectroscopy

Jablonski Diagram

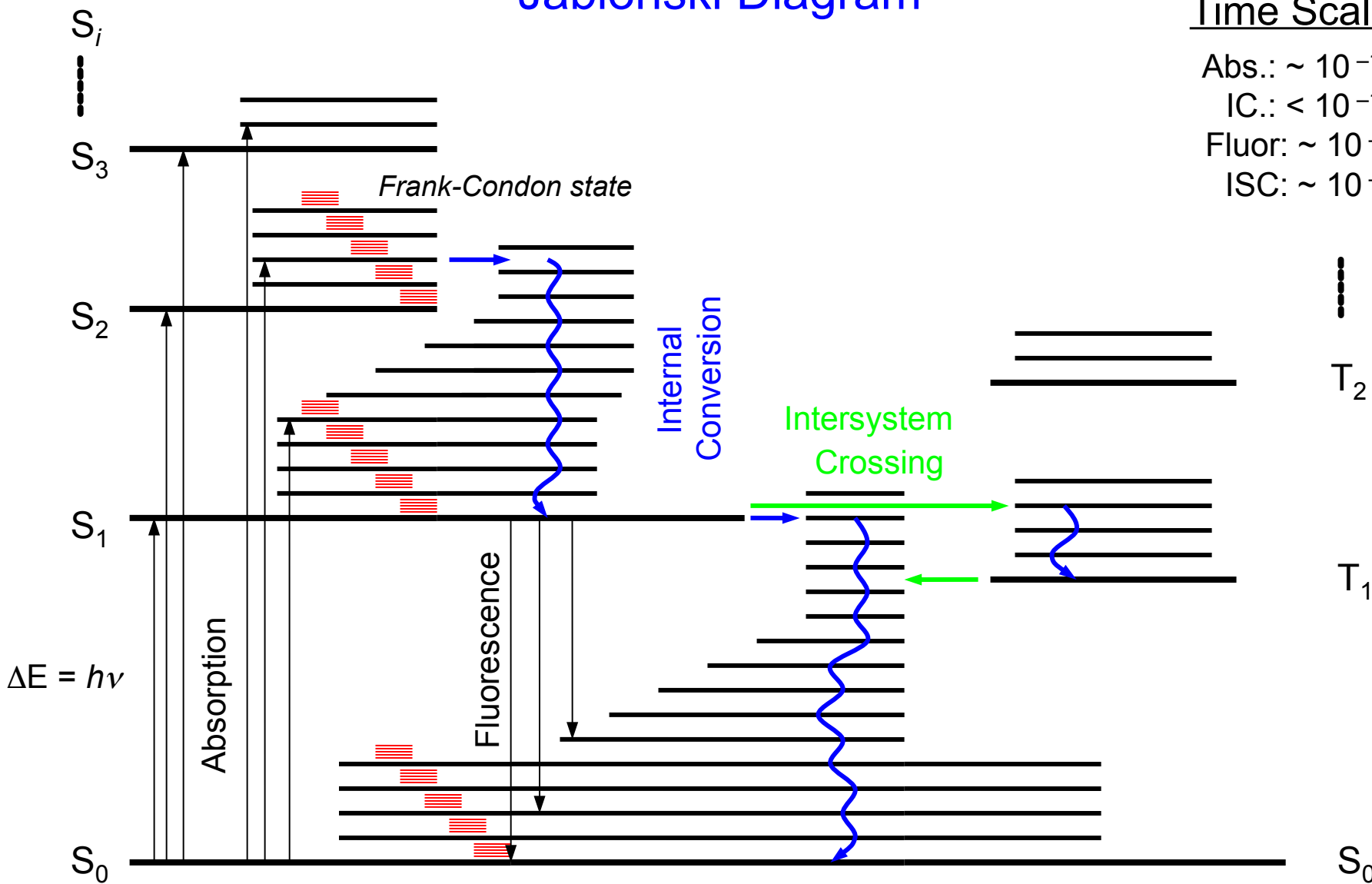
Time Scales

Abs.: $\sim 10^{-15}$ s

IC.: $< 10^{-12}$ s

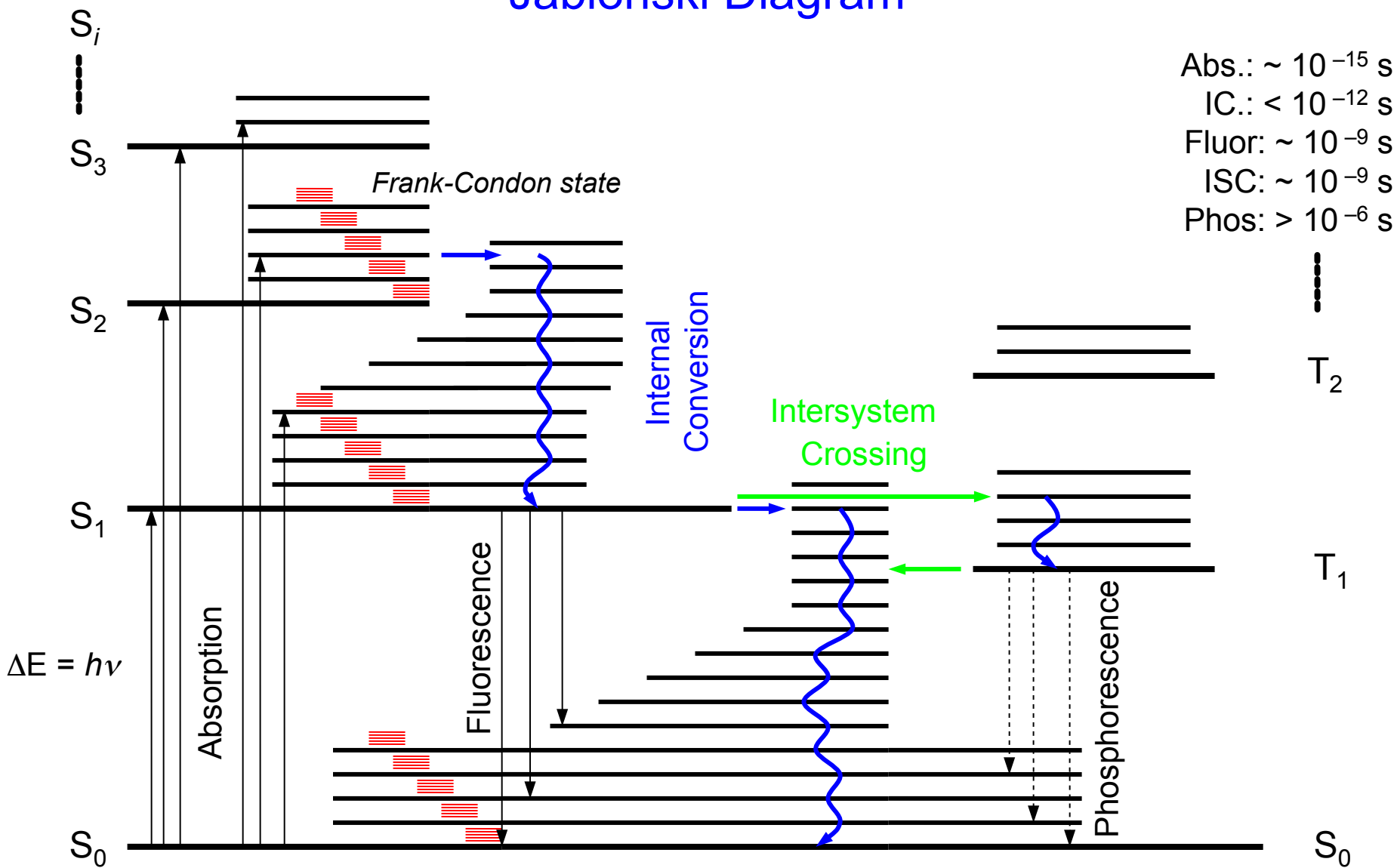
Fluor.: $\sim 10^{-9}$ s

ISC: $\sim 10^{-9}$ s



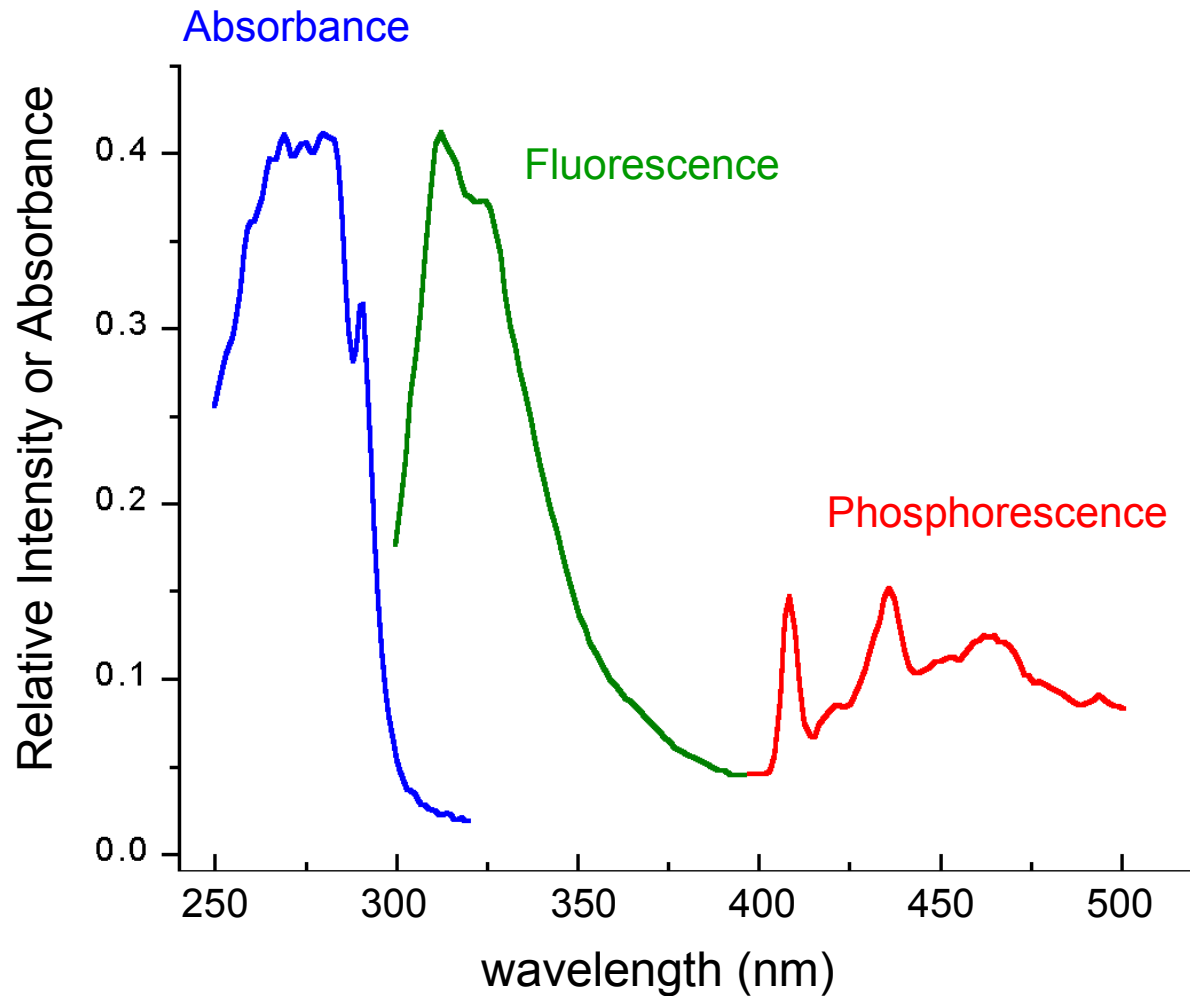
Spectroscopy

Jablonski Diagram



Spectroscopy

Single Trp Residue in Cod Parvalbumin; 77 K



Spectroscopy

Loss of Energy from Excited State back to Ground State

- Internal Conversion (IC; mainly through vibrational relaxation)
- Quenching: collisions with solvent, solutes, or groups of chromophore
- Intersystem Crossing (ISC)
 phosphorescence from long-lived triplet state
- Förster Resonance Energy Transfer (FRET)
- Emission of a photon
 fluorescence from lower energy than from initial Frank-Condon state;
 Stoke's shift
- Excited-State Reactions